

The Dose Effect of Whey Protein on Glycemic Control in Adults with Insulin Resistance

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Abstract: Whey protein coupled with a glucose challenge increases insulin secretion and may decrease glucose responses in people with pre-diabetes and type 2 diabetes. These responses may be attributed to whey protein's effect on the incretins glucagon-like peptide-1 (GLP-1) and glucose dependent insulinotropic polypeptide (GIP). The purpose of this study was to examine the effect of various doses of whey protein on postprandial glycemic control and incretin responses. Participants with insulin resistance ($n=9$, mean \pm SD; age: 64.3 ± 8.1 yrs; BMI: 29.4 ± 6.0 kg/m²; fasting plasma glucose: 6.9 ± 1.2 mmol/l; HbA1c: $6.4 \pm 0.6\%$) completed three randomized treatments. Treatment 1 included 250 ml water + 20 g whey protein (T₁), and treatment 2 included 250 ml water + 30 g whey protein (T₂). The control treatment included 250 ml water (CON). Each treatment was followed by a 50 g oral glucose tolerance test. Incremental area under the curve (iAUC) for insulin increased from CON to T₁ ($P<0.01$, 45.5%), CON to T₂ ($P<0.01$, 61.0%), and T₁ to T₂ ($P<0.01$, 28.5%), with a significant decrease in postprandial AUC for glucose with T₂ ($P=0.04$, -41.2%). Neither GIP nor GLP-1 iAUC increased with T₁ or T₂ compared to CON. However, postprandial glucose iAUC was significantly reduced for T₂ compared to CON ($P=0.04$, -41.2%). There was a dose effect of whey protein on plasma insulin with a significant decrease in postprandial glucose iAUC following T₂. Thirty grams of a whey protein preload may be adequate to provide postprandial glycemic improvements in the disease management of type 2 diabetes or pre-diabetes

Keywords: Diabetes, Pre-diabetes, GIP, GLP-1, Glucose, Glucagon

1. Introduction

Insulin resistance (IR) is defined as an impairment in glucose homeostasis resulting from decreased insulin sensitivity in multiple body tissues including skeletal muscle and liver [1]. IR leads to increased blood glucose concentrations, resulting in many individuals having developed pre-diabetes or type 2 diabetes. There are many strategies used to treat IR including a combination of medications, diet, exercise, and weight loss [2]. Incretin-

based therapies are also commonly prescribed to patients with IR to decrease chronically high blood glucose and high levels of glycated hemoglobin (HbA1c) [3]. The incretin hormones glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1) account for approximately 50–70% of insulin secretion following nutrient ingestion in healthy individuals [4]; however, the incretin response following nutrient intake is reduced in people with type 2 diabetes [5]. Recent investigations have implemented whey protein as a potential treatment for the

management of IR [6, 7]. Whey protein augments insulin secretion through an increase of the incretin hormones in people with type 2 diabetes [8], while subsequently decreasing blood glucose concentrations in both healthy [9], and type 2 diabetes [10].

Decreases in glucose area under the curve (AUC) are evident when 50 g of whey protein is consumed 30 min prior to nutrient ingestion [10]. Recent literature has suggested smaller doses of whey protein such as 15 g for those with type 2 diabetes [7] and 20 g for obese males [6], consumed prior to meal ingestion can reduce postprandial glucose. In healthy individuals, there is a dose dependent postprandial increase in insulin, with a subsequent decrease in glucose following whey protein ingestion of 5-20 g [11], 10-40 g [9], and 10-20 g [12]. However, it is unknown if there is a similar dose effect of whey protein in individuals with IR. Therefore, the purpose of this study was to examine the effect of 20 g (T₁) and 30 g (T₂) of whey protein isolate ingested 30 min prior to a 50 g oral glucose tolerance test (OGTT) on plasma glucose, insulin, C-peptide, GIP, GLP-1 and glucagon responses in individuals with IR. Our hypothesis was that both T₁ and T₂ would result in an increase in insulin secretion and a decrease in plasma glucose concentrations, with T₂ producing the greatest improvements.

2. Materials and Methods

2.1. Study Population

Nine sedentary male participants with IR were recruited to participate in this study ($n=9$, mean \pm SD; age: 64.3 \pm 8.1 yrs; BMI: 29.4 \pm 6.0 kg/m²; fasting plasma glucose: 6.9 \pm 1.2 mmol/l; HbA1c: 6.4 \pm 0.6%). Sedentary was defined by <150 min of physical activity per week. Participants taking any medications known to affect rate of gastric emptying or hypoglycemic agents other than sulfonylurea or metformin were excluded. Participants completed a preliminary blood draw that was analyzed for fasting plasma glucose and HbA1c to determine qualification for the study based on American Diabetes Association guidelines [13]. Participants with a fasting plasma glucose > 5.56 mmol or HbA1c > 5.7% were selected. This study was approved by the Texas Woman's University Institutional Review Board.

2.2. Anthropometrics

Body composition was measured using dual-energy x-ray absorptiometry (DXA; Lunar Prodigy; GE Healthcare, Madison, WI). Height, weight, and body mass index (BMI) were determined. Height was measured using a stadiometer (Perspective Enterprises; Kalamazoo, MI) and weight was measured to the nearest 0.1 kg using a digital scale (Tanita Corp.; Arlington Heights, IL).

2.3. Study Design

Participants completed three randomized treatments that consisted of consuming: 250 ml of water (CON), 250 ml of water + 20 g whey protein (T₁), and 250 ml of water + 30 g

whey protein (T₂), followed by a 50g OGTT. The whey protein (Isopure Whey Protein Isolate, Nature's Best, Hauppauge, New York, United States) and CON were consumed as a preload (30 min prior to the 50 g OGTT) and treatments were separated by a minimum of seven days. Participants recorded a three-day diet log preceding each treatment, with the dinner meal the night prior to each OGTT kept consistent for macronutrient composition and timing.

2.4. Oral Glucose Tolerance Test (OGTT)

Participants arrived at the laboratory between 0500 and 0700 hrs following a 10 hr fast to complete a 50g OGTT. A venous catheter was placed in an antecubital vein and the participant rested for 5 min before the first blood draw (baseline). A 0.9% saline solution with a 1/5s drip rate was used to keep the line patent. Following the first blood draw, participants consumed a designated whey protein preload corresponding to T₁, T₂, or CON. Another blood draw was taken 30 min after the preload, which was immediately followed by the 50g OGTT. Additional samples were collected at 15, 30, 60, 90, 120, and 150 min after the consumption of the OGTT beverage (Trutol Dextrose, ThermoFisher Scientific, Waltham, United States). Blood samples were collected into 4 ml EDTA tubes containing 1.25 Pefabloc/ml of blood (Roche Diagnostics, Mannheim, Germany), 5 μ l Protease Inhibitor/ml of blood (EMD Millipore Corporation, Billerica, Massachusetts, United States), and 5 μ l Protease Cocktail/ml of blood, and Protease Cocktail/ml of blood (EMD Millipore Corporation, Billerica, Massachusetts, United States). Blood samples were immediately centrifuged at 3000 rpm for 10 min at 10°C. Plasma was aliquoted into cryogenic vials and stored at -80°C until analysis. Samples thawed for one hour prior to plasma analysis of glucose, insulin, C-peptide, GIP, GLP-1 and glucagon. Plasma glucose was analyzed using a YSI 2900D glucose analyzer (Yellow Spring Inc, Yellow Springs, Ohio, United States). Plasma hormone concentrations of C-peptide, GIP, GLP-1, insulin, and glucagon were analyzed using a Luminex Human Metabolic Hormone multiplex assay (HMHEMAG-34, EMD Millipore, Billerica, MA). Twenty-five μ l of each sample were analyzed in duplicate.

2.5. Statistical Analysis

Incremental area under the curve (Δ AUC) was calculated using the trapezoidal method. Two areas were calculated, Δ AUC (baseline to 150) and postprandial Δ AUC (0-150) to analyze the responses of whey protein and the OGTT alone. Repeated-measures analysis of variance (RM ANOVA) was used to determine significant differences in Δ AUC for all dependent variables. The differences at each time-point across treatments were also analyzed by RM ANOVA. Statistical significance was set at $P<.05$. If results were significant, a Bonferroni post-hoc test was used for pairwise comparisons. The statistical analysis for glucagon was ($n=6$) due to low values outside the range of the kit.

3. Results

3.1. Participant Characteristics

A total of nine participants completed the study. Table 1 displays participant descriptive characteristics. There were no significant differences in baseline fasting values between the three treatments (see Figure 3).

Table 1. Participant Characteristics.

Age (y)	64.3	8.1
Height (m)	1.68	0.1
Weight (kg)	83.21	20.3
BMI (kg/m ²)	29.41	6.0
Body Fat (%)	42.51	7.8
HbA1c (mmol/mol)	45.8	4.3
HbA1c (%)	6.37	0.6
Fasting Plasma Glucose (mmol)	6.85	1.2

Data are presented as mean \pm SD.

3.2. Incremental AUC

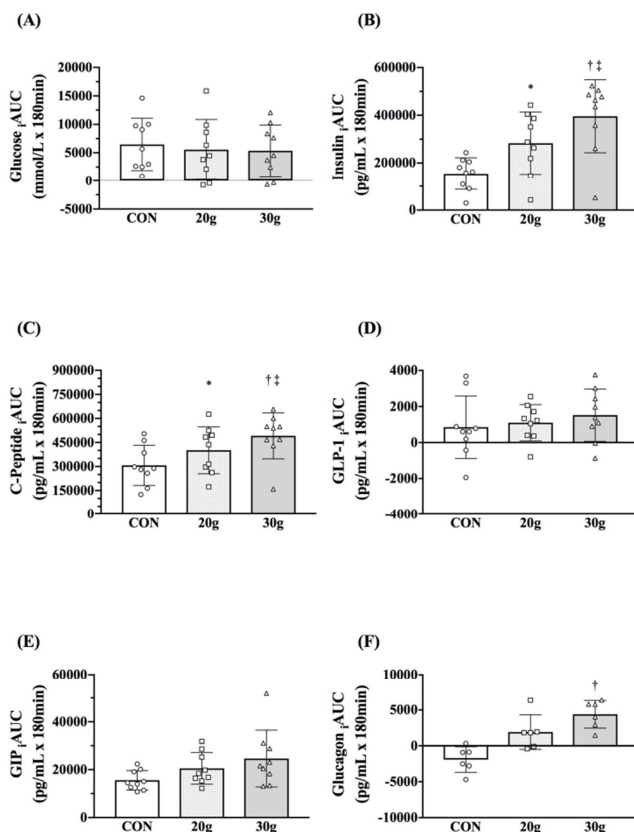


Figure 1. $iAUC$ of CON, T_1 , and T_2 . For every time-point, plasma was analyzed for glucose (A), insulin (B), C-peptide (C), GLP-1 (D), GIP (E), and glucagon (F). * represents significant differences for T_1 compared to CON. † represents significant differences for T_2 compared to CON. Circles: CON, squares: T_1 , triangles: T_2 . ‡ represents significant differences for T_2 compared to T_1 .

Glucose $iAUC$ was not different between the 3 treatments (Figure 1A). Insulin $iAUC$ (Figure 1B) was significantly greater in T_1 compared to CON ($P<0.01$, 45.5%), T_2 compared to CON ($P<0.01$, 61.0%), and the T_2 compared to

T_1 ($P<0.01$, 28.5%). C-peptide $iAUC$ (Figure 1C) was greater in T_1 compared to CON ($P<0.01$, 23.5%) and T_2 compared to CON ($P<0.01$, 37.5%). Glucagon $iAUC$ (Figure 1F) was greater in T_2 compared to CON ($P<0.01$, 143%). There were no significant differences in $iAUC$ for total GLP-1 (Figure 1D) or GIP (Figure 1E).

3.3. Postprandial $iAUC$

Glucose postprandial $iAUC$ (Figure 2A) was significantly reduced following T_2 compared to CON ($P=0.04$, -41.2%) and insulin postprandial $iAUC$ (Figure 2B) was significantly higher for T_2 compared to CON ($P=0.042$, 38%). GIP postprandial $iAUC$ (Figure 2E) was significantly decreased for T_1 compared to CON ($P=0.021$, -27.2%), and T_2 compared to CON ($P=0.01$, -20.4%) with no significant changes for GLP-1 across all treatments. The glucagon postprandial $iAUC$ (Figure 2F) was lower for both T_1 compared to CON ($P=0.01$, -79.9%), and T_2 compared to CON ($P=0.032$, -76.0%).

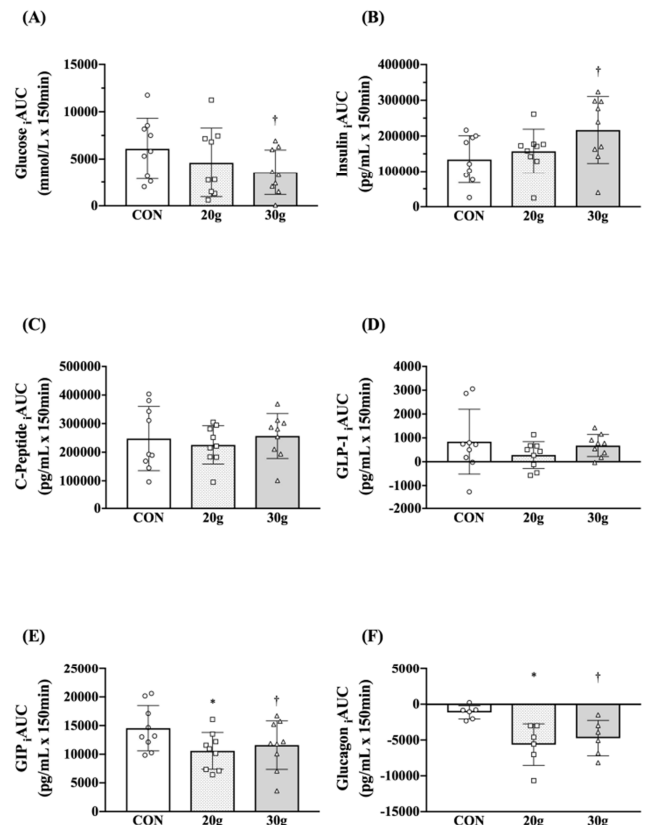


Figure 2. Postprandial $iAUC$ for CON, T_1 , and T_2 . Calculations were made for glucose (A), insulin (B), C-peptide (C), GLP-1 (D), GIP (E), and glucagon (F). * represents significant differences for T_1 compared to CON. † represents significant differences for T_2 compared to CON.

3.4. Time-point

Glucose (Figure 3A) was not different between treatments for any time-points. Insulin (Figure 3B) was significantly increased at min 30 for T_2 compared to CON ($P<0.01$) and was continuously increased through min 90. C-peptide (Figure 3C) was elevated in a similar manner as insulin

without significant differences between treatments. GLP-1 (Figure 3D) increased following T₁ compared to CON at min 60 and min 120 ($P=0.02$; $P=0.02$). GLP-1 was also elevated for T₂ at min 90 and 120 compared to CON ($P<0.01$, $P<0.01$). GIP (Figure 3E) was not different between treatments for any time-points. Glucagon (Figure 3F) increased following T₁ and T₂ consumption compared to CON ($P=0.04$, $P=0.03$) respectively. Glucagon remained elevated for T₂ compared to CON through min 90 before returning to baseline values.

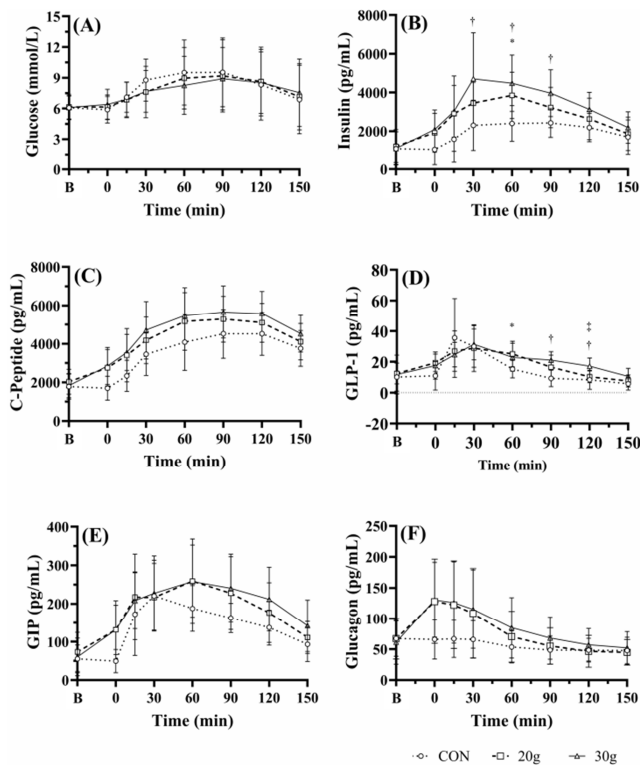


Figure 3. Treatments of CON, T₁, and T₂. For every time-point, plasma was analyzed for glucose (A), insulin (B), C-peptide (C), GLP-1 (D), GIP (E), and glucagon (F). * represents significant differences for T₁ compared to CON. † represents significant differences for T₂ compared to CON. Circles: CON, squares: T₁, triangles: T₂. ‡ represents significant differences for T₂ compared to T₁.

4. Discussion

In the current experiment, there was no decrease in glucose iAUC ; however, postprandial iAUC for glucose was significantly decreased following T₂ of whey preload as shown in Figure 2A. Previous literature has reported decreases in glucose concentration following similar doses of whey protein in obese males and those with insulin resistance [6, 7]. One study examining a 20g whey protein 15 min preload observed decreases in postprandial glucose concentrations [6]. King and colleagues also observed a decrease in postprandial glucose AUC following 15g of whey consumed with a mixed meal in men with IR [7]. Whey protein quantities of 40 g and higher have also led to increased insulin and decreased plasma glucose responses in individuals with type 2 diabetes [10, 14] and prediabetes

[15]. Jakubowicz et al. (2014) reported a decrease in both peak glucose and glucose 180 min AUC following the ingestion of 50 g of whey protein mixed with 250 ml of water as a preload, prior to a high glycemic breakfast in individuals with type 2 diabetes [8]. Although the current study did not examine changes in peak glucose for T₂, there was an equivalent increase in plasma insulin using a similar research design as Jakubowicz and colleagues with a decrease in glucose postprandial iAUC [8].

The decrease in postprandial iAUC for glucose can be attributed to the increase in insulin concentration 30 minutes following T₁ and T₂, prior to glucose ingestion (Figure 2B). Insulin and C-peptide complete iAUC (Figure 1B, 1C) appear to have a concomitant rise with T₁ and T₂ whey protein ingestion. Approximately 50-70% of insulin secretion is stimulated by incretins [4]. Although whey protein has been reported to stimulate release of GLP-1 and GIP, the current study did not observe changes in complete iAUC (Figure 1D, 1E). Conversely, postprandial iAUC for GIP was reduced following T₁ and T₂ of whey protein ingestion (Figure 2E). However, as shown in Figure 3E, postprandial iAUC for GIP did in fact increase immediately following the ingestion of whey. Individuals with insulin resistance are reported to have an impaired incretin effect [16]. This could explain the absence of a rise in GLP-1 (Figure 1D) and GIP (Figure 1E).

Insulin secretion may also be increased directly through amino acids. Certain positively charged amino acids are suggested to potentiate insulin secretion through an increase in depolarization of pancreatic beta cells [17, 18]. Arginine stimulates a calcium influx that depolarize the voltage gated channels, leading to insulin secretion [17]. L-arginine is known to have a similar effect by increasing intracellular calcium uptake, resulting in insulin secretion [18]. Studies have also reported branched-chain amino acids such as leucine and isoleucine may have a direct effect on insulin secretion through pancreatic beta cells [19, 20]. Another potential reason for the increase in insulin without a subsequent decrease in plasma glucose concentration could be the inhibition of glucose uptake in skeletal muscle. Protein ingestion has been suggested to induce skeletal muscle insulin resistance in mice by inhibiting AKT via the mTOR pathway [21]. The current study did not examine skeletal muscle activity.

Lastly, Figure 1F illustrates an acute increase in glucagon following the ingestion of whey protein. Insulin suppresses hepatic glucose production by more than 50%, while also inhibiting glucagon [22, 23]. Glucagon is also inhibited by GLP-1. Conversely, glucagon is enhanced through increases in GIP [24]. Based on Figure 1D and 1E, GLP-1 or GIP do not seem to have an influence on the increased glucagon complete iAUC (Figure 1F). This is a similar result to Frid et al. where no changes in GLP-1 following 27.6 g of whey consumption in individuals with type 2 diabetes were observed [25]. However, Frid and colleagues did observe an increase in GIP iAUC [25]. A decrease in postprandial iAUC for glucagon was observed in the present study (Figure 2F), but this was due to the initial rise following the whey protein preload. Based on the complete iAUC for glucose (Figure 1A), insulin (Figure 1B), and

glucagon (Figure 1F), the authors speculate glucagon may play a stronger role than expected in increasing plasma glucose than insulin has on lowering plasma glucose.

5. Conclusion

The current study found that whey protein doses of 20 g and 30 g administered 30 min prior to an OGTT resulted in a concomitant rise in plasma insulin complete iAUC ; however, plasma glucose was unaffected. Previous literature suggests 40 g and 50 g of whey protein can provide glycemic benefit in individuals with IR [9, 10]. Contradictory to previous studies examining lower doses, the results of the current study suggest that 20 g and 30 g of whey protein may not be beneficial in individuals with IR. Whey protein may provide benefits in healthy individuals but there may other mechanisms inhibiting the effectiveness for individuals with IR. Neither T_1 nor T_2 of whey protein may be adequate to provide glycemic improvement in the disease management of pre-diabetes or type 2 diabetes. More research is needed to provide a greater understanding of the effects whey protein has on individuals with IR. An advantage of consuming whey preload is delaying gastric emptying in individuals with type 2 diabetes [10]. In the current study, whey protein consumed as a preload does not appear to be the most ideal strategy. However, as shown with postprandial iAUC for glucose, this technique appeared to have beneficial results from the preprandial rise in insulin following T_2 of whey consumption.

Limitations

The sample size of the present study was too small to differentiate between individuals with pre-diabetes ($n=4$) and type 2 diabetes ($n=5$). However, both groups appeared to have similar results. Another limitation is the use of only two separate doses of whey protein. Previous literature has suggested larger doses are beneficial in the population recruited for this study. The novelty of this study was in determining if lower doses were more effective.

Acknowledgements

TC, CI, SD, and VB designed the study. TC, CI, RG, MB, and SD recruited participants and collected data. TC, CI, RG, and AH analyzed the blood samples. TC and MB analyzed the data. TC, CI, RG, MB, SD, MS and VB drafted the manuscript. All of the authors approved the final version of the manuscript.

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